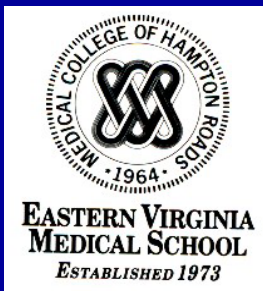


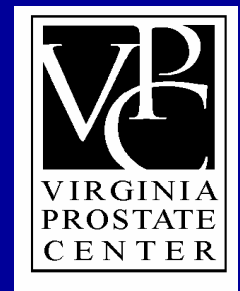
Signal and Noise: Garbage in Garbage out

O. John Semmes

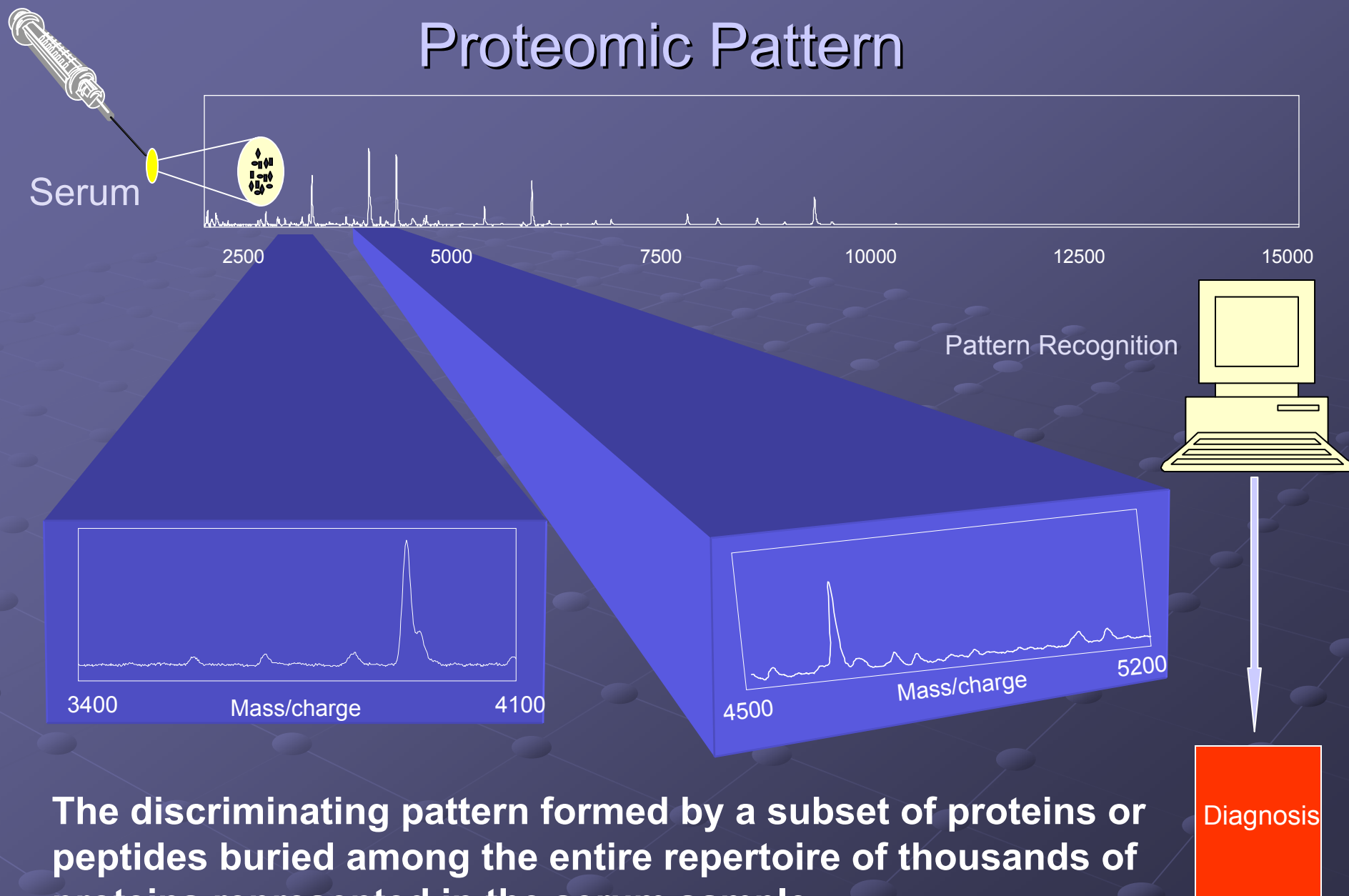
***Director, Center for Biomedical Proteomics
Department of Microbiology and Molecular Cell Biology
Virginia Prostate Center
Eastern Virginia Medical School Norfolk, VA***



*Eastern Virginia Medical School
Discovery Laboratory*



Proteomic Pattern



The discriminating pattern formed by a subset of proteins or peptides buried among the entire repertoire of thousands of proteins represented in the serum sample.

THINGS WE CAN DO TO MAKE THE PATTERNS BETTER

- 1. Some things to do before the Mass Spec (TOF-tube)**
- 2. Some things to do after the Mass Spec (TOF-tube)**

Achieving Reproducibility in SELDI

1. Get a gatekeeper for experimental design and interpretation.
2. Use “standard” samples; external (spiked) proteins, internal proteins, serum sample for QC.
3. Synchronization/optimization of instrument output using the QC sera. Laser/detector settings.
4. Constant monitoring and adjustment of parameters.
5. Automation of sample processing steps.
6. Find out what the peaks are! (robustness)

Table 1. QC spectra criteria

Protein	Signal to Noise Ratio (S/N)	Resolution
Insulin	N/A	600
IgG	700	N/A
Peak1: 5906 \pm 0.2%	>40	>400
Peak 2: 7768 \pm 0.2%	>80	>400
Peak 3: 9289 \pm 0.2%	>80	>400

Quality control assessment of the PBSII is based on signal to noise ratio (s/n) and resolution. The table provides the s/n and resolution required for each QC protein used in Phase IA for a site to proceed to Phase II.

QC standardization At six Validation Sites

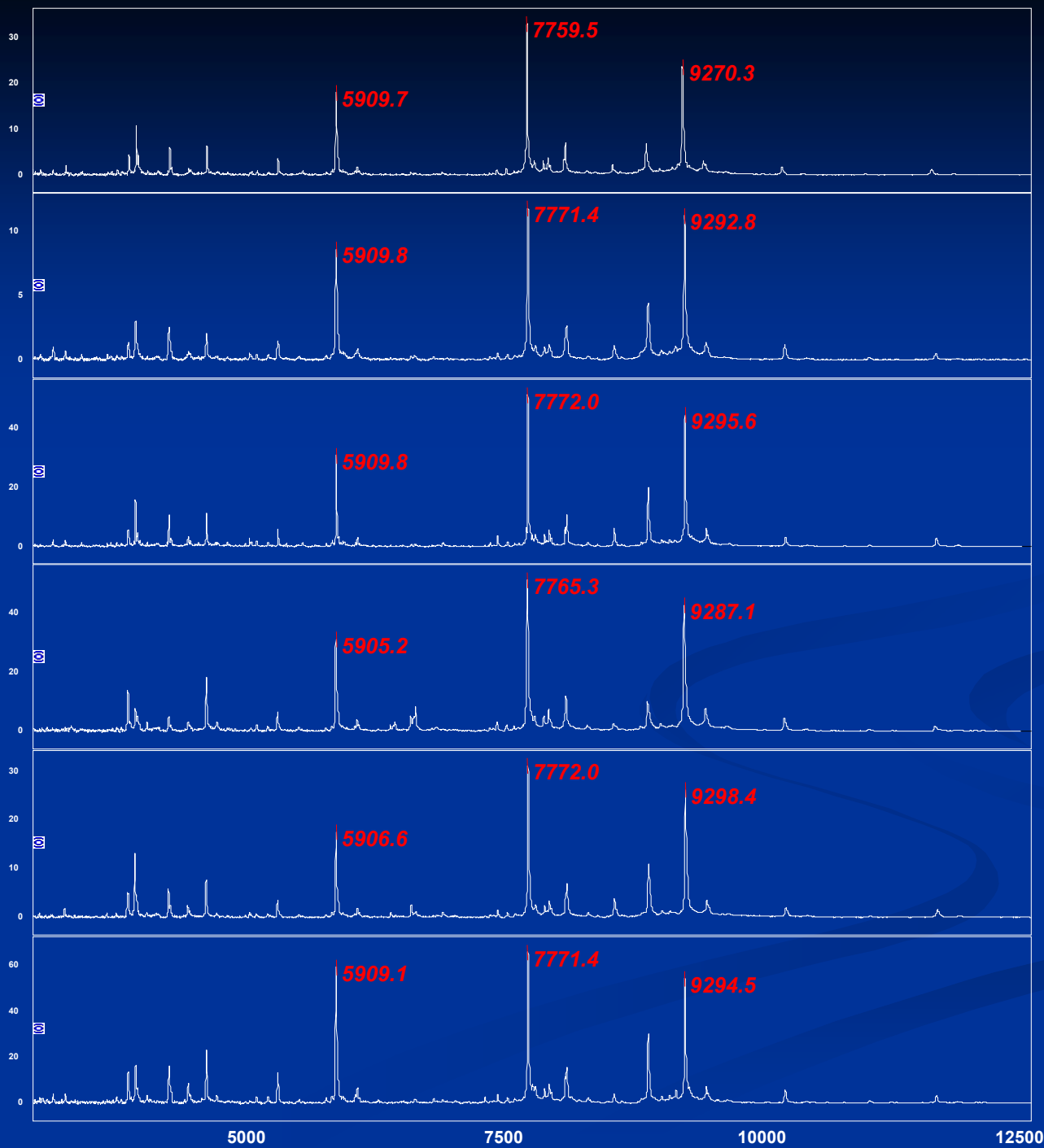
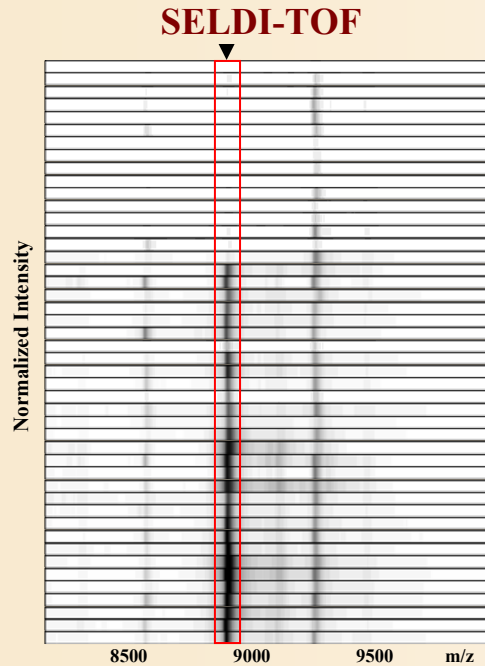


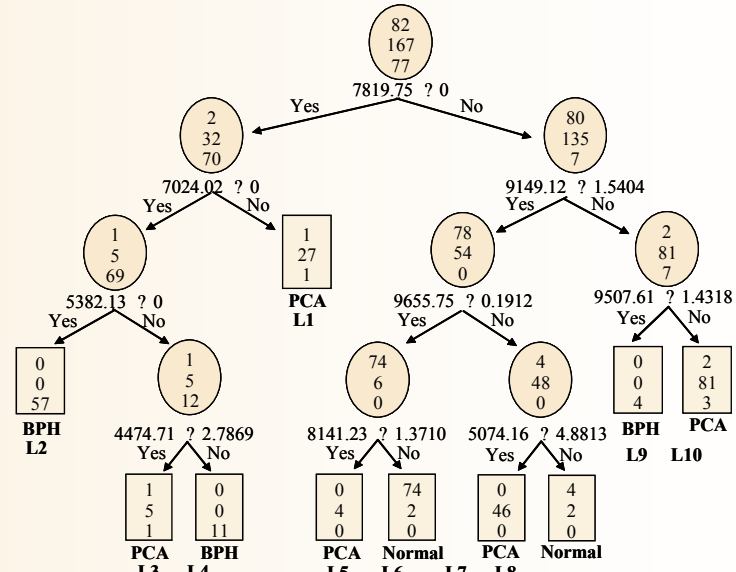
Table 2b Inter-Lab variability

		Mass	Intensity	S/N	Resolution
<i>Peak 1</i>	average	5906.47	26.57	163.06	460.73
	stdev	6.70	9.67		107.72
	CV	0.0011	0.36		0.23
<i>Peak 2</i>	average	7768.61	35.94	242.75	505.54
	stdev	8.41	6.25		82.77
	CV	0.0010	0.17		0.16
<i>Peak 3</i>	average	9289.18	30.96	244.03	439.28
	stdev	9.89	4.70		77.35
	CV	0.0011	0.15		0.18

Summary of Biomarker Discovery and Identification

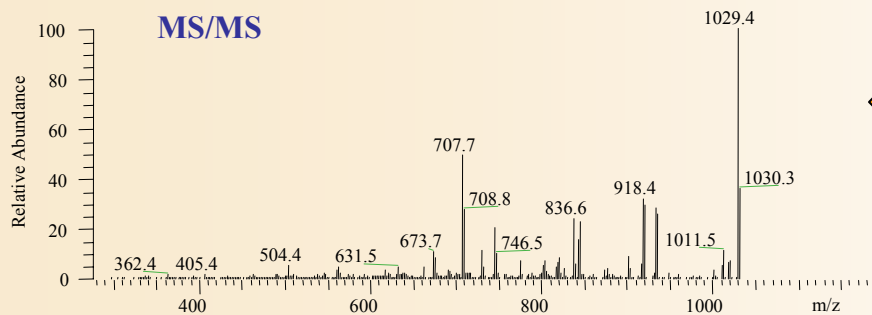


Classification and Regression Tree Analysis

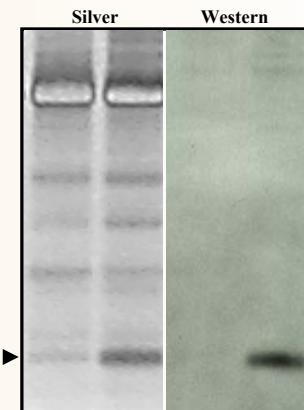


[Cancer Research 62, 3609-3614, July 1, 2002] © 2002
[American Association for Cancer Research](http://www.aacr.org/)

Identification



Purification



Things to do after the tube

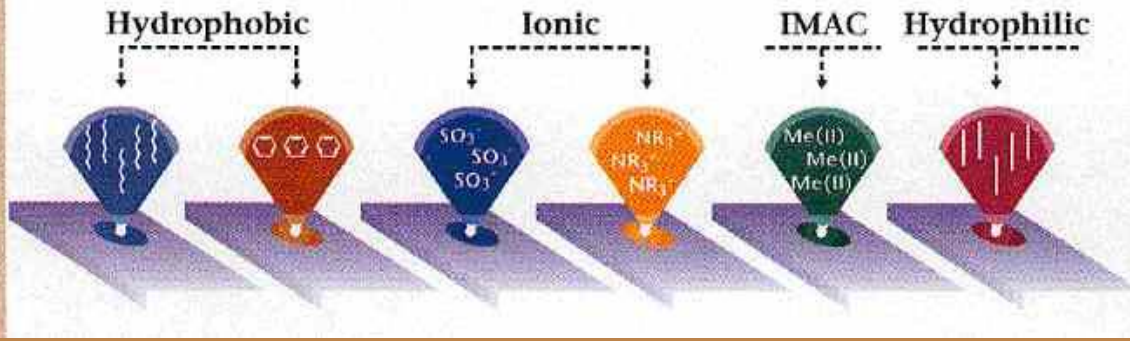
Analysis of Source of Variation

1. Metrologic Analysis of SELDI-TOF Process.
2. Spectral Analysis of Output.

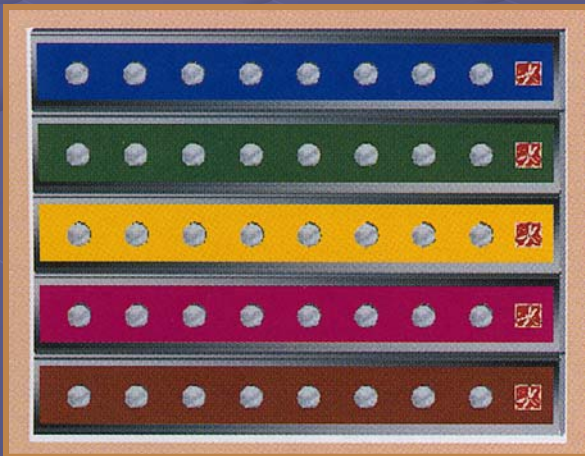
Proteomics Using SELDI Technology

Surface Enhanced Laser Desorption

Chemical Surfaces



← **Surface Chemistries** Each chip binds a specific set of proteins based on the chromatographic surface of the ProteinChip®.



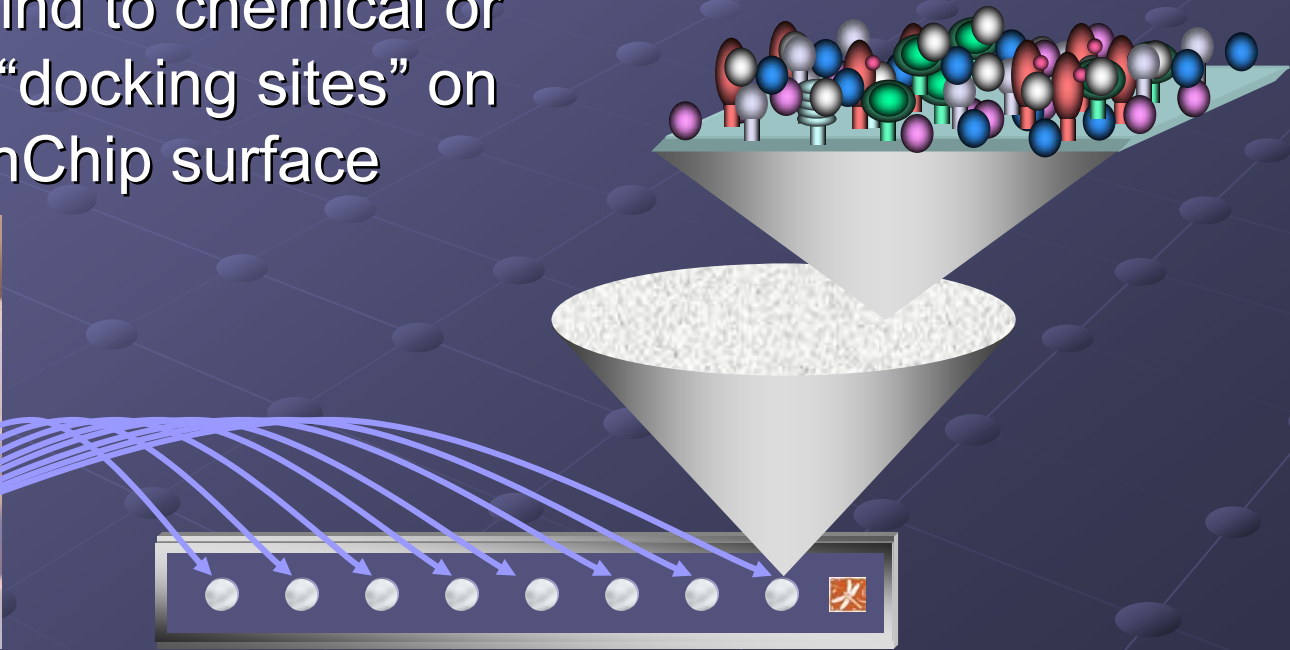
← Protein Chips

Each spot on the chip will contain sample from a control or diseased/treated source. The spots are analyzed separately and a mass spectra is created for each spot representing the proteins bound to the chip surface.



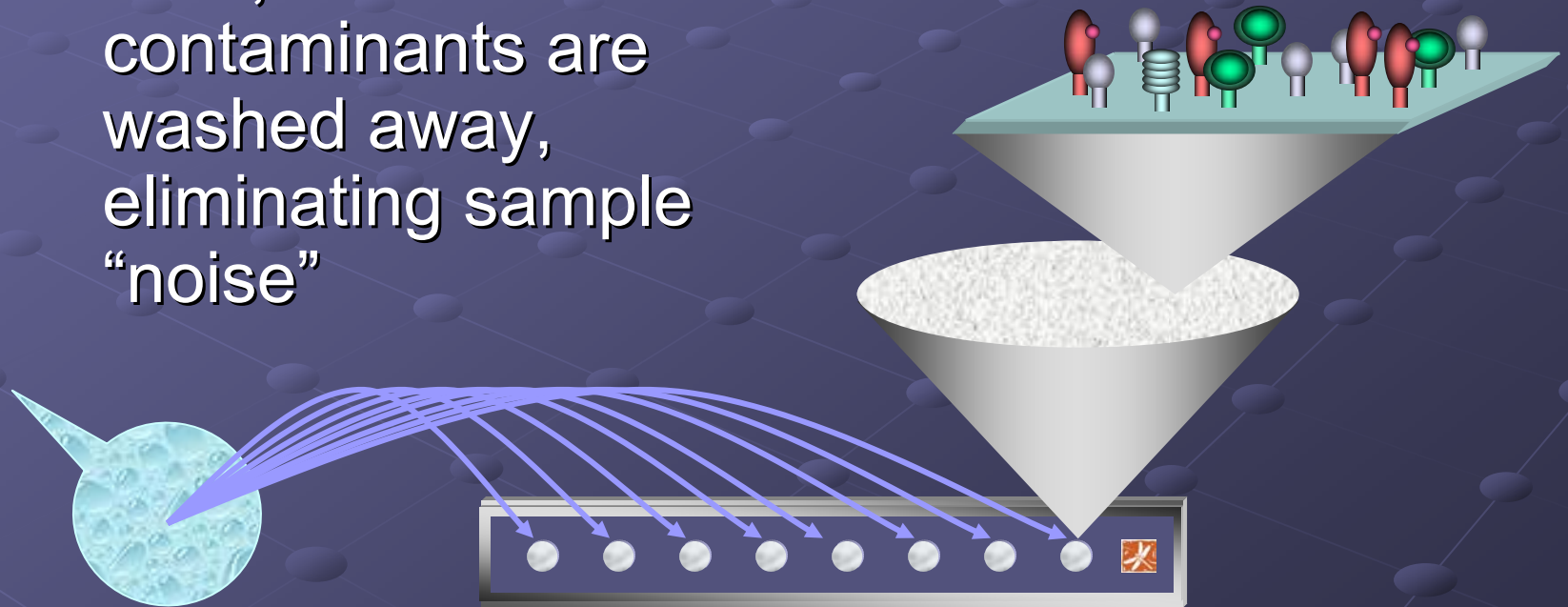
ProteinChip Technology: Protein Binding

- Crude sample is placed (and processed) on a ProteinChip Array
- Proteins bind to chemical or biological “docking sites” on the ProteinChip surface



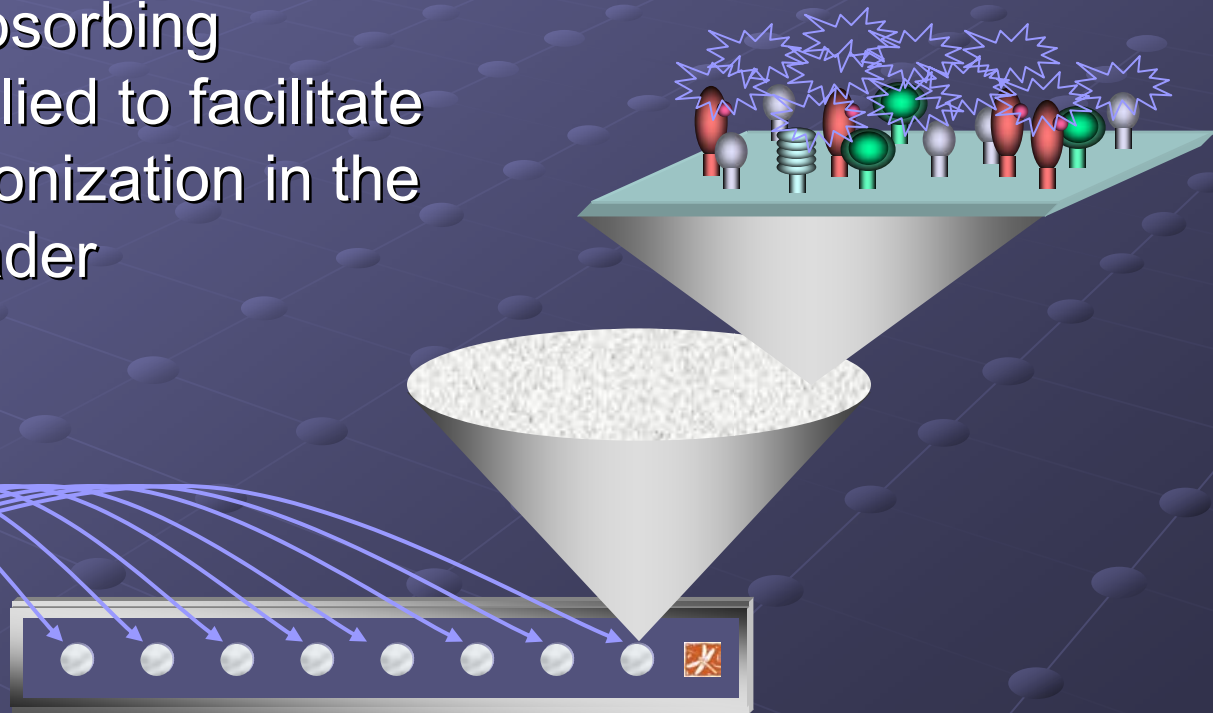
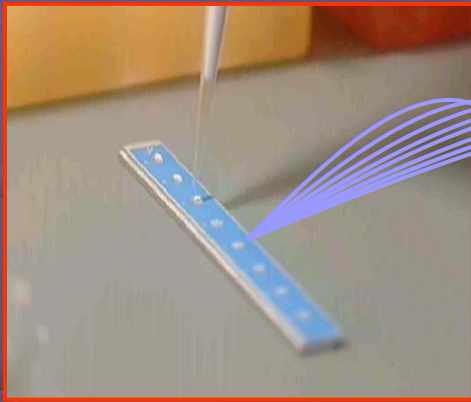
ProteinChip Technology: Washing Reduces Non-Specific Binding

- Non-binding proteins, salts, and other contaminants are washed away, eliminating sample “noise”

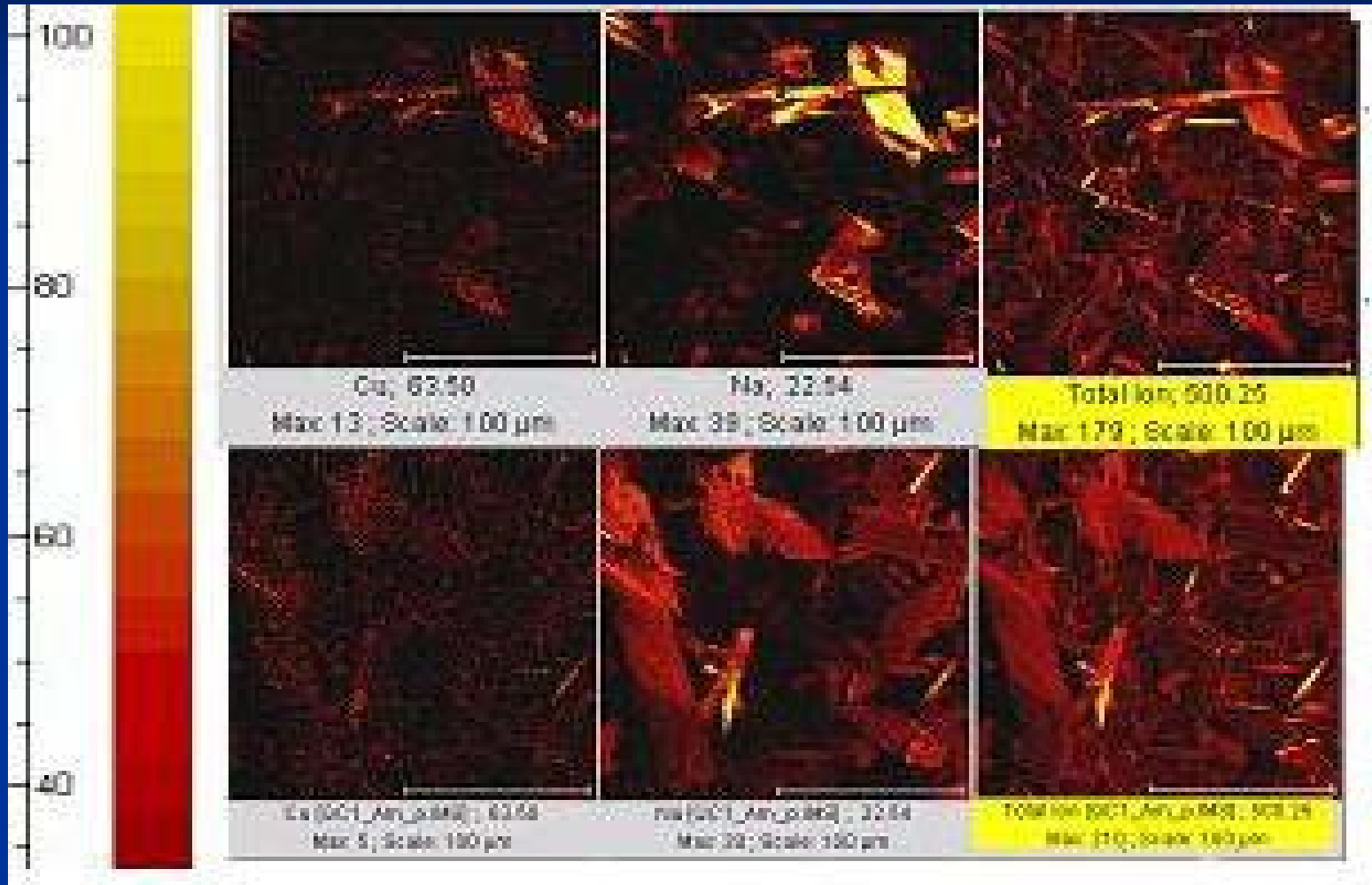


ProteinChip Technology: Addition of EAM

- EAM (Energy Absorbing Molecule) is applied to facilitate desorption and ionization in the ProteinChip Reader



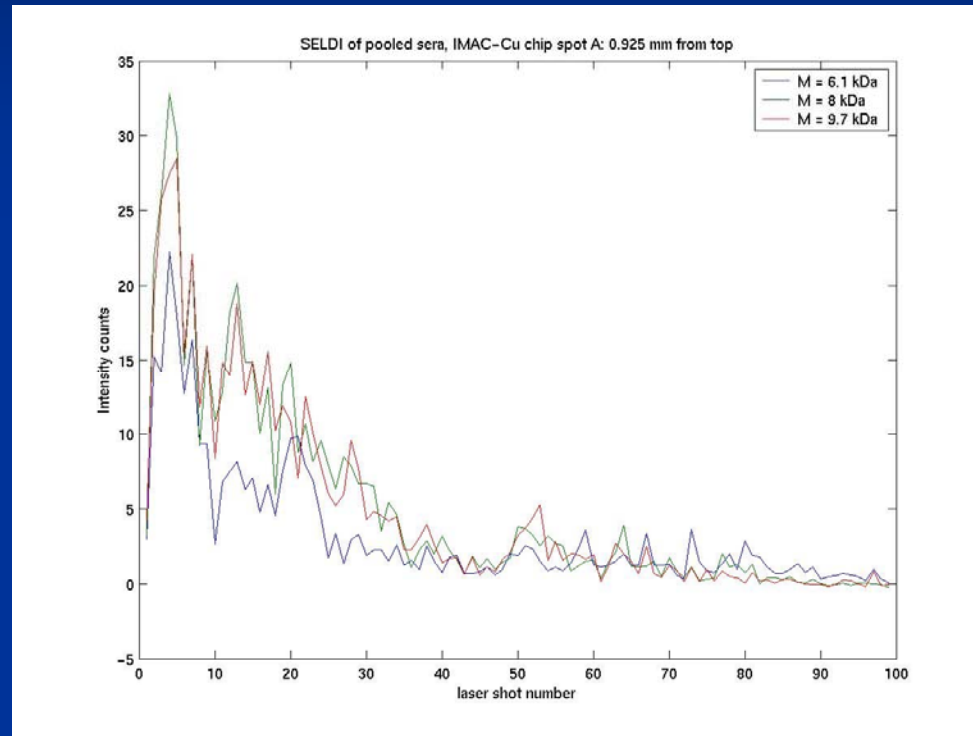
Desorption Surface



-

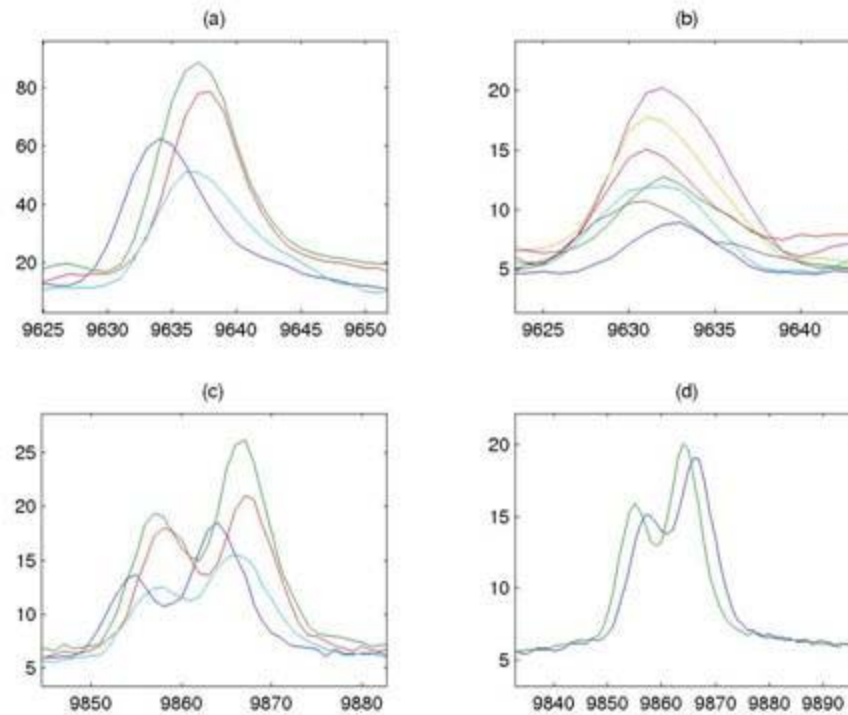
+

Limited Inefficient Desorption

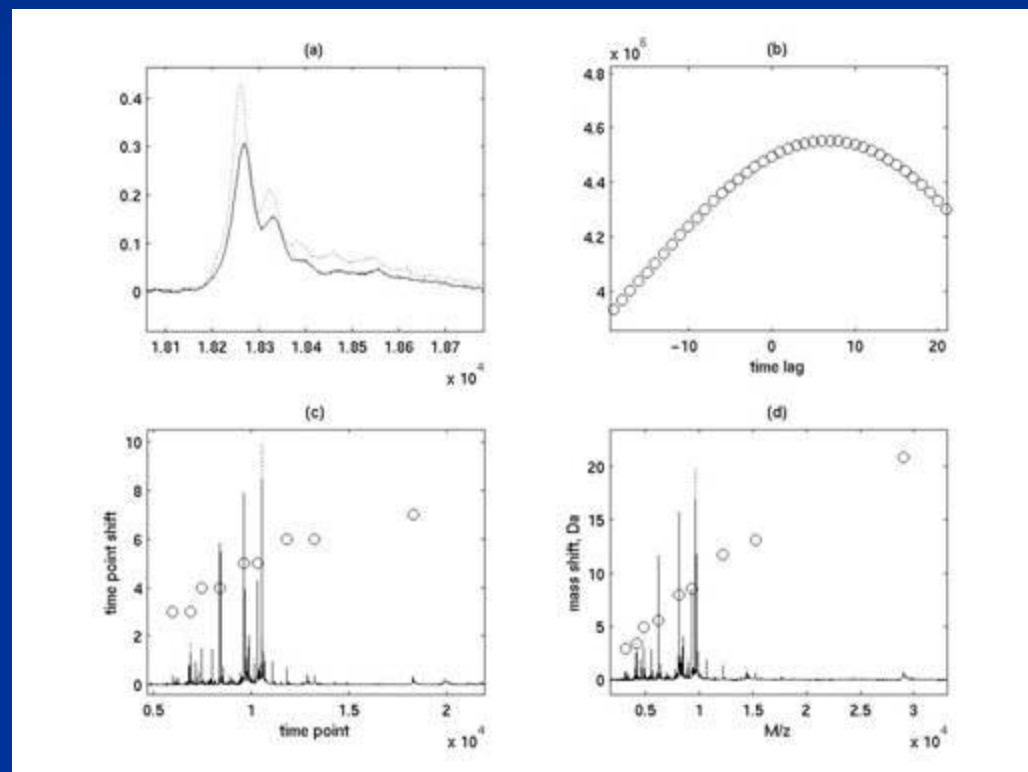


Nano-Scale Surface Polishing

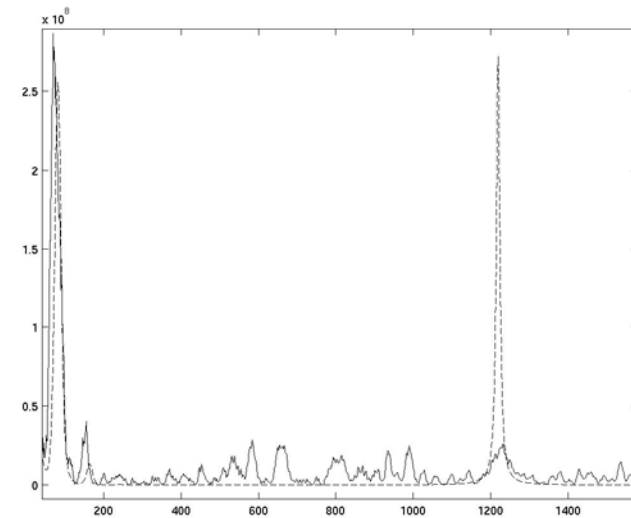
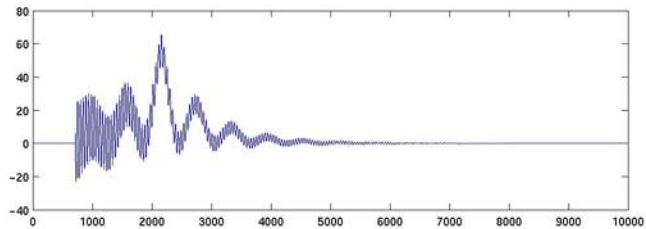
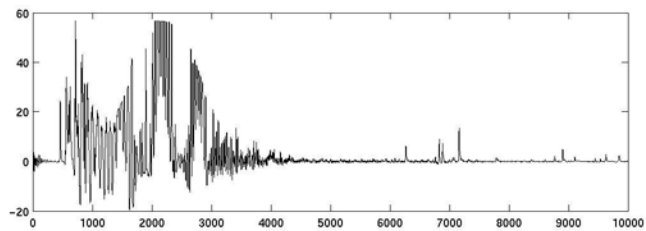
Peak Jitter Between Single Laser Shots Reduced Resolution



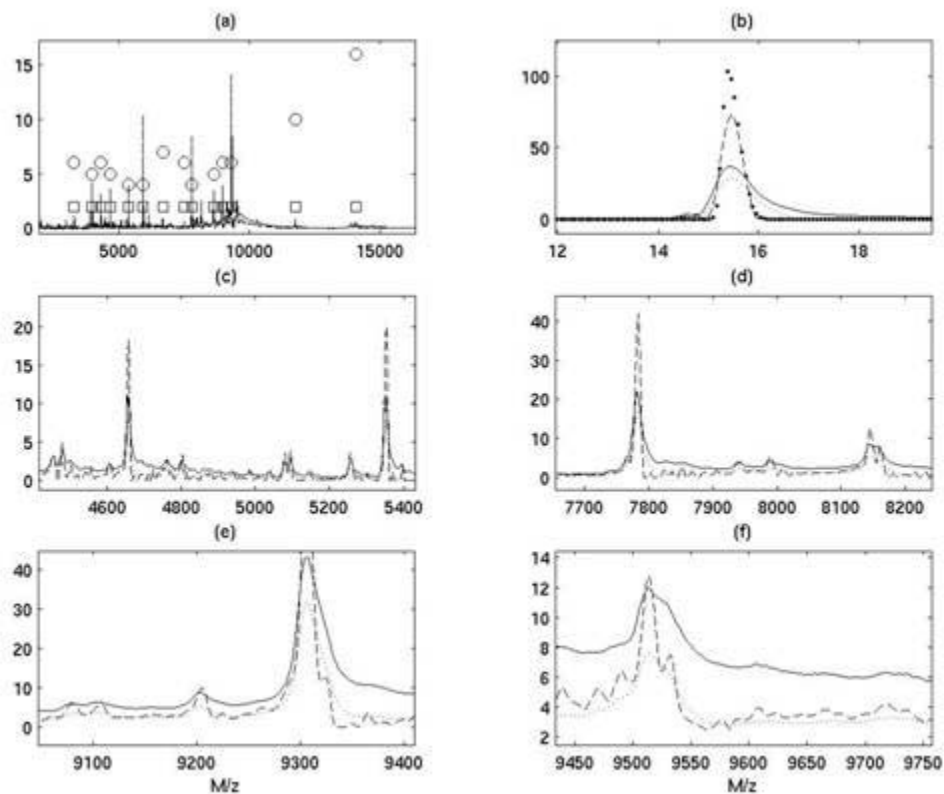
Automatic Dejitter



Denoising Filters

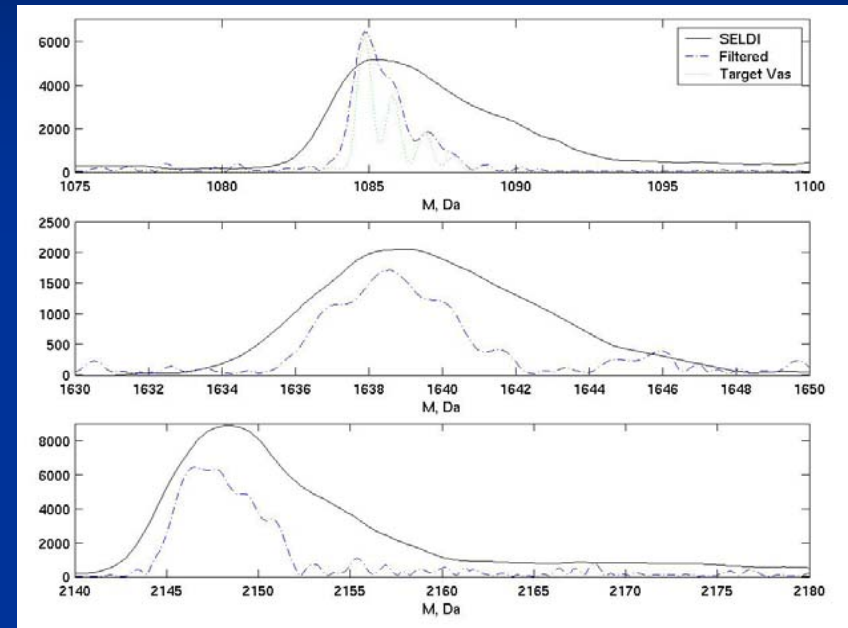
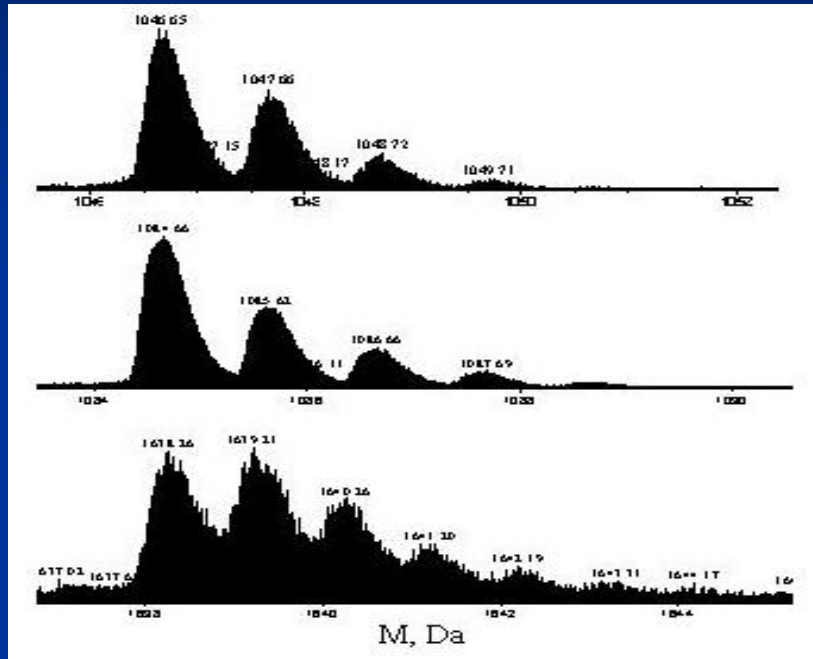


Trace Add-Back Filters



Model based target filtering

Trace Add-Back Filters



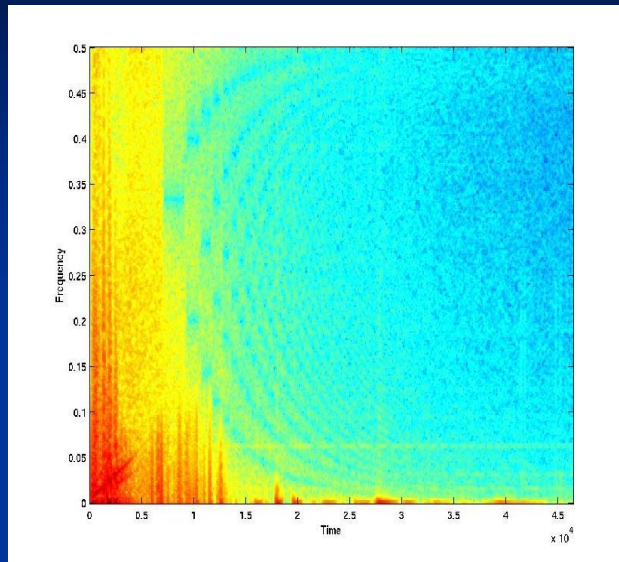
Peptide standards with SIMS-resolved isotopic structure

Best approach may involve
Internal Standards with known isotope structure

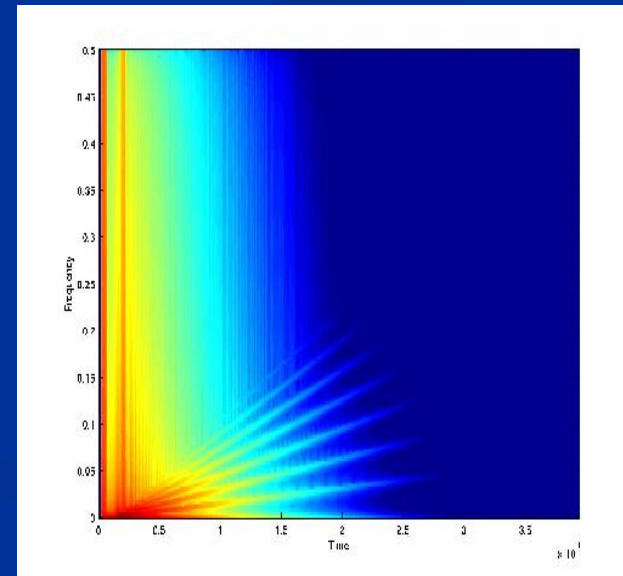
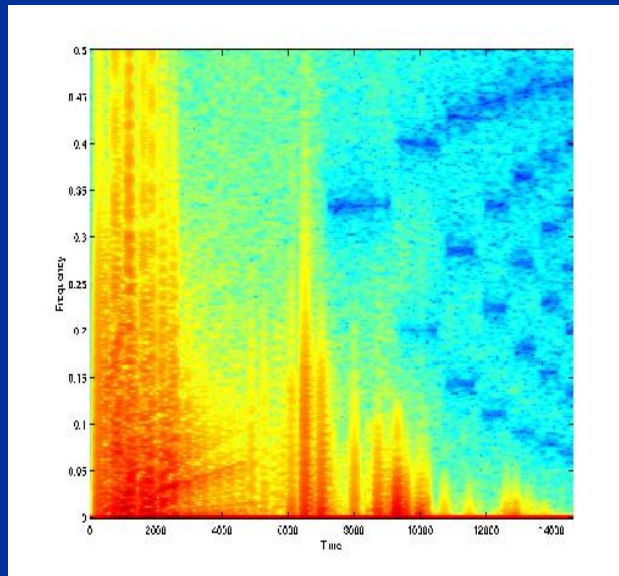
Placing external proteins in data valleys

The success of denoising filters depends on defining baseline

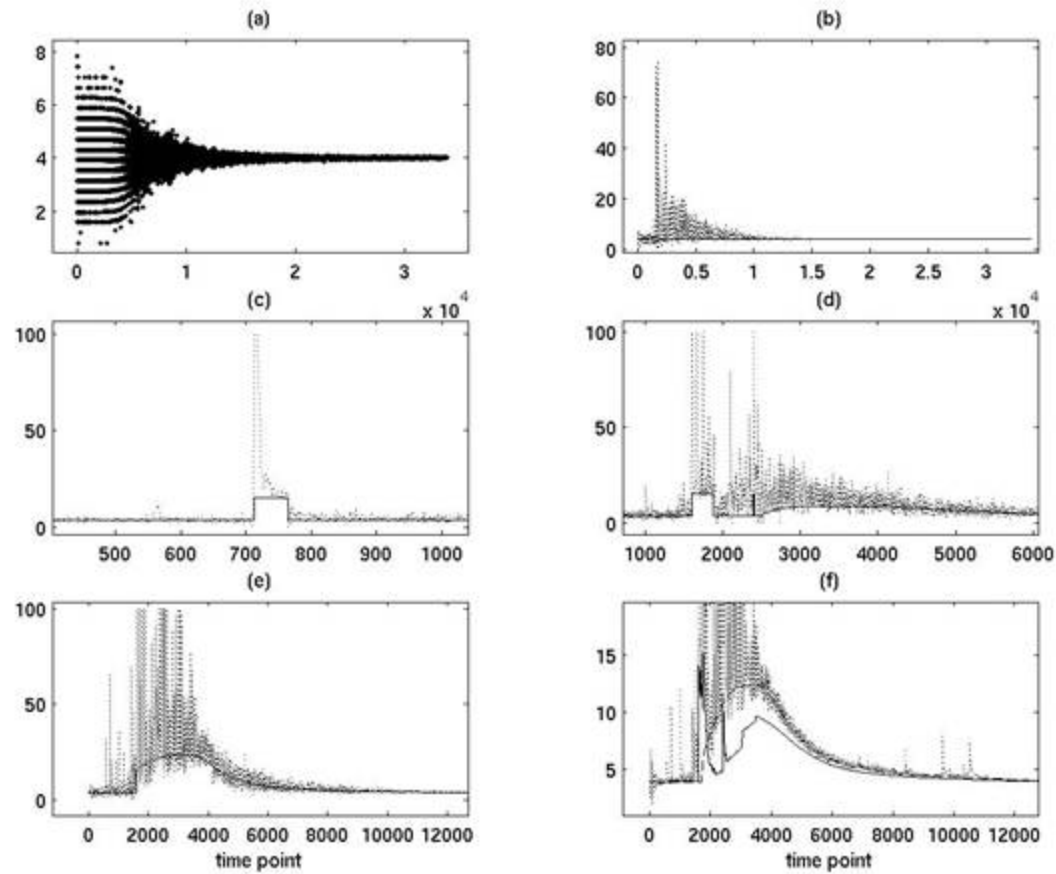
Spectral Analysis



Detector Overload

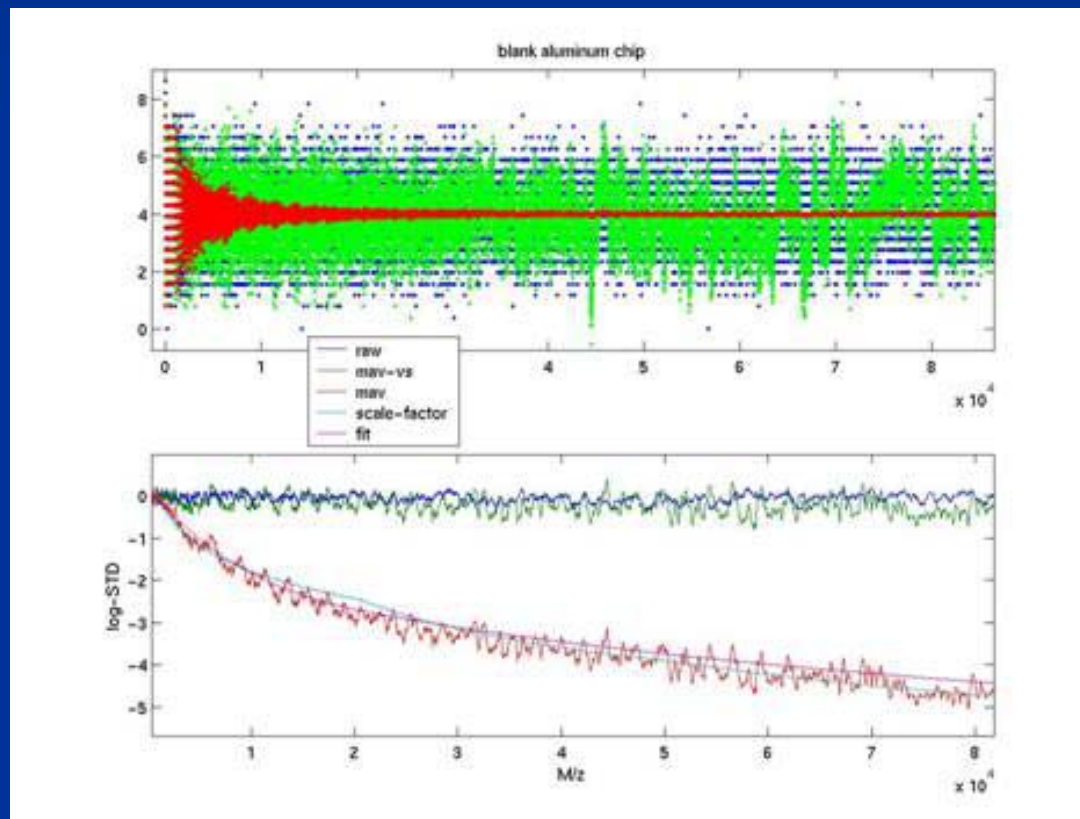


Effect of Detector Overload On Baseline

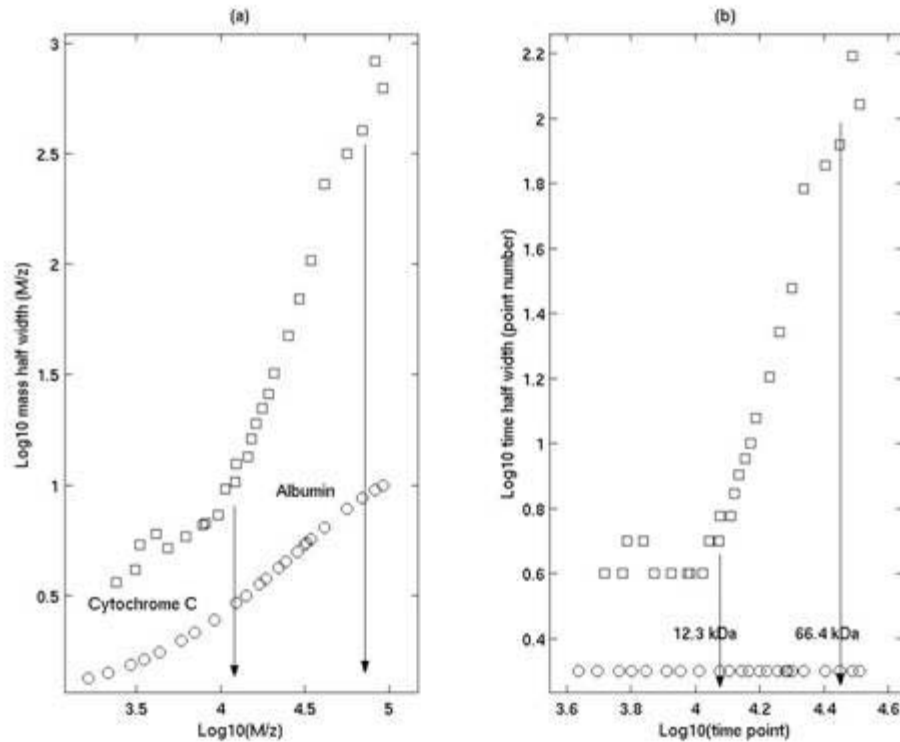


Variance Rescaling

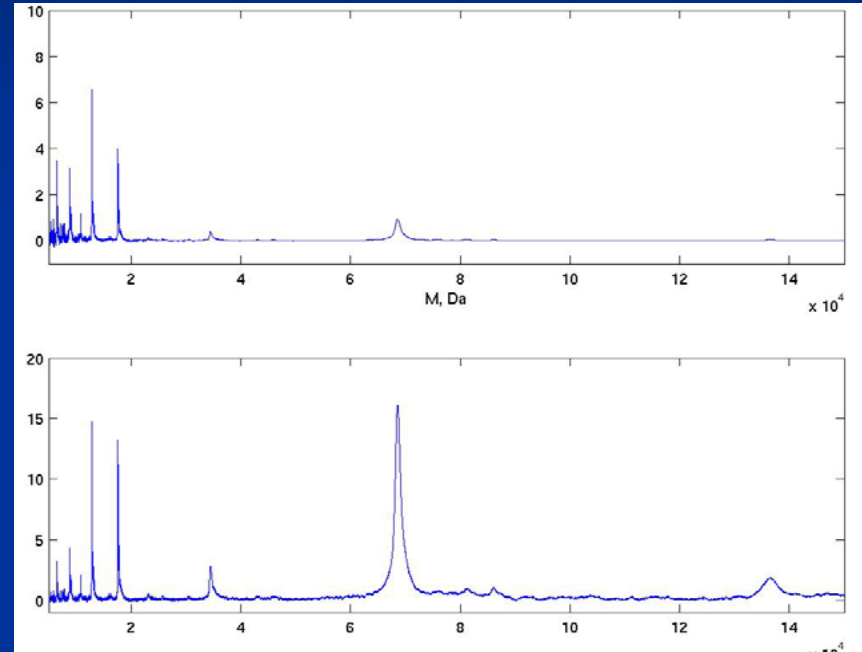
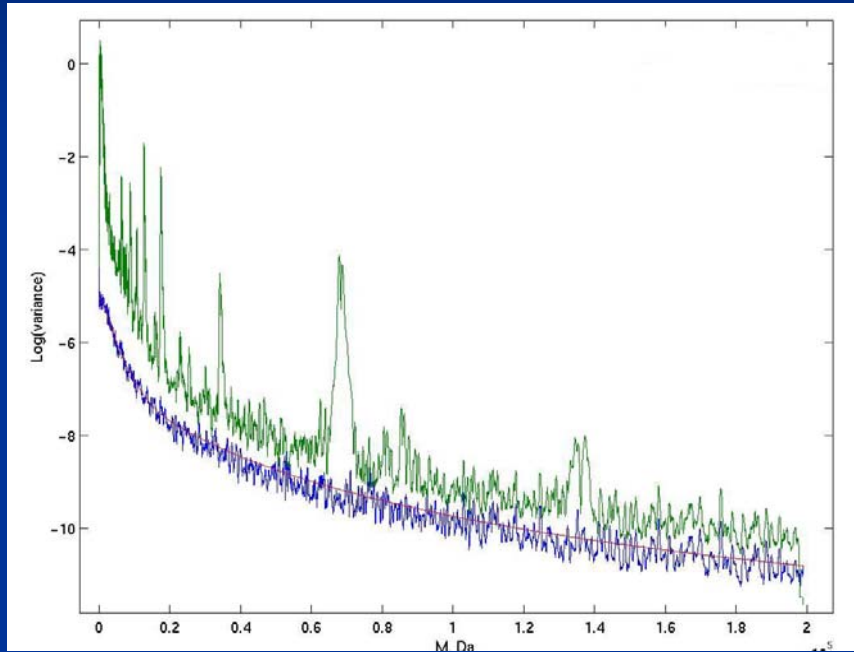
Effect of Default Moving Average Filter



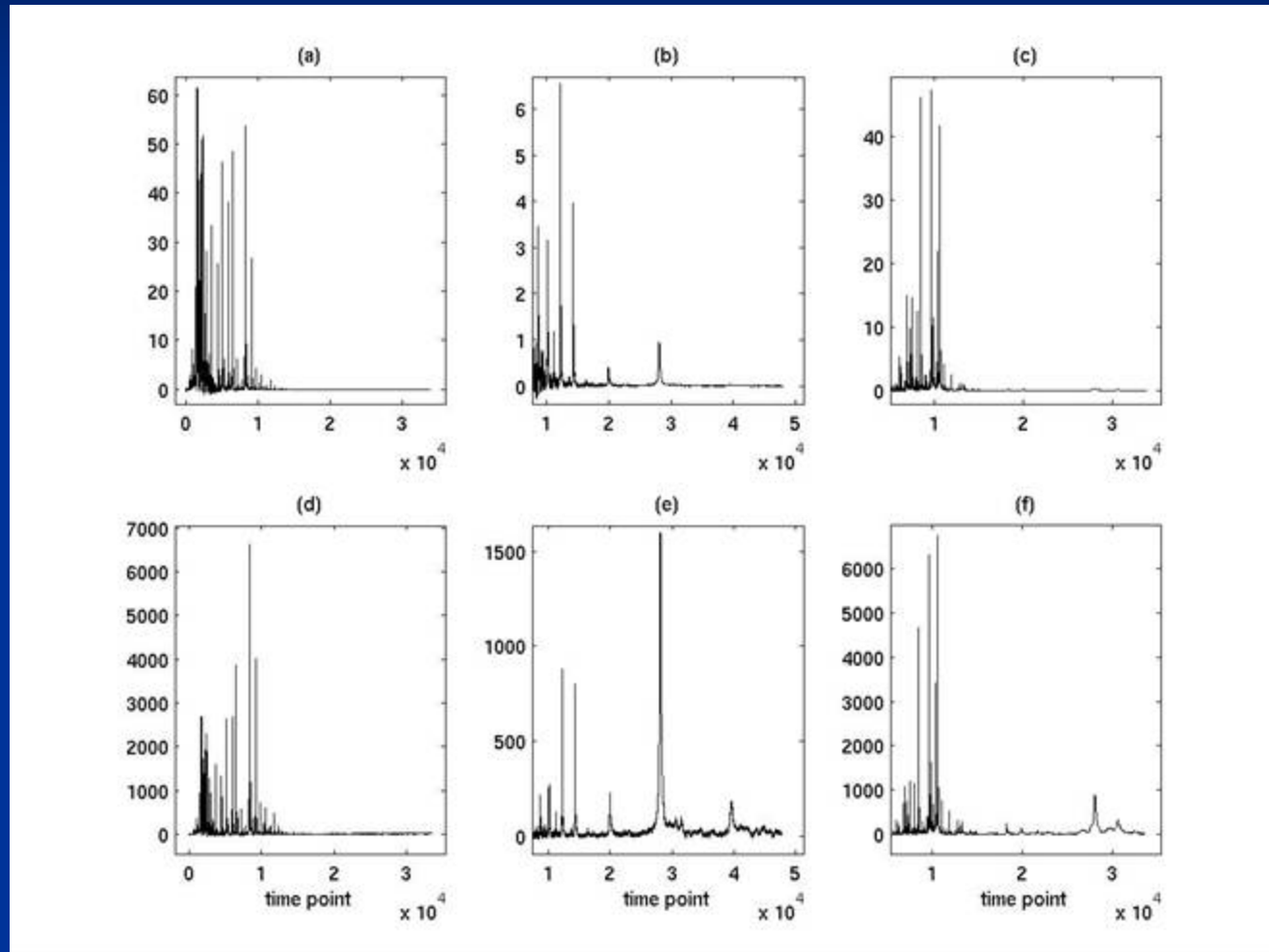
Mass dependence of peak width and default MAV



Mass Dependence of Variance



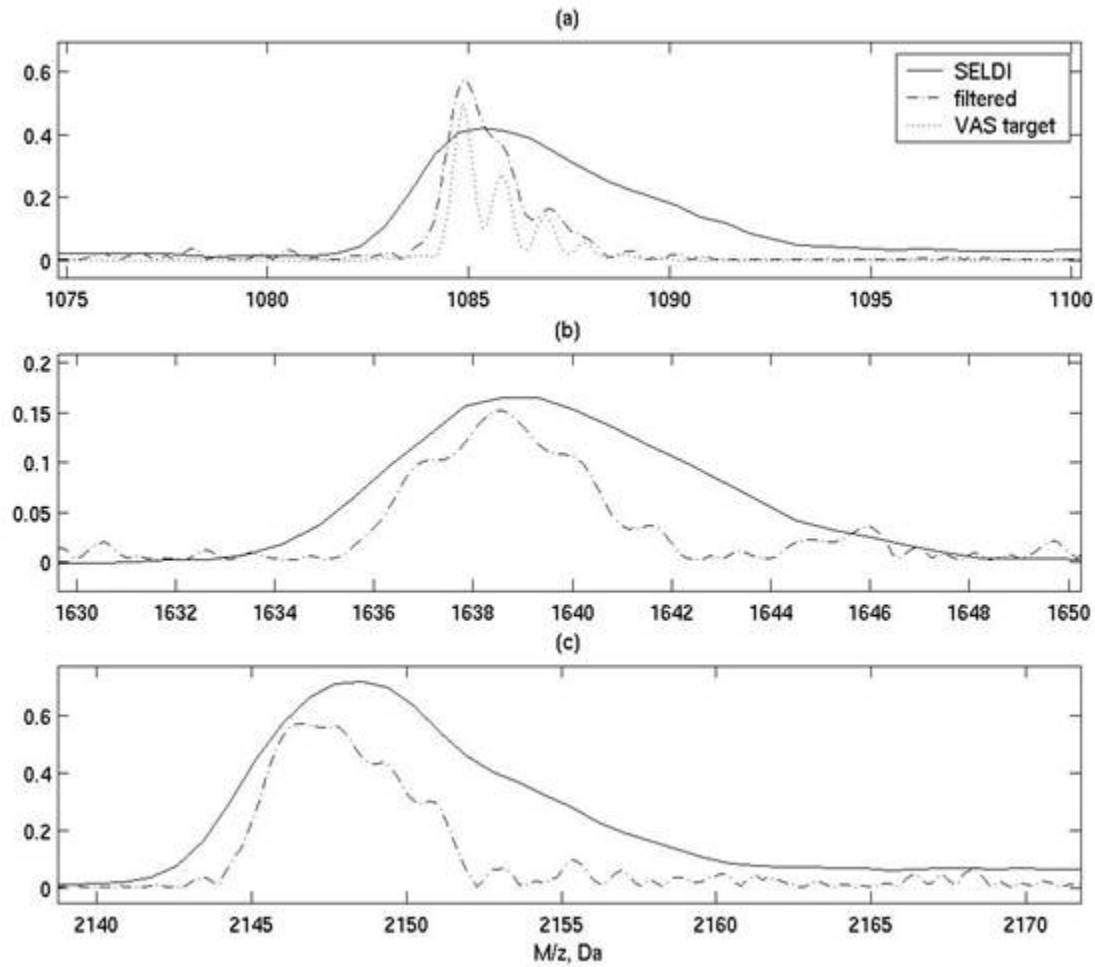
Variance Rescaling: Stationary Noise, Increased Sensitivity



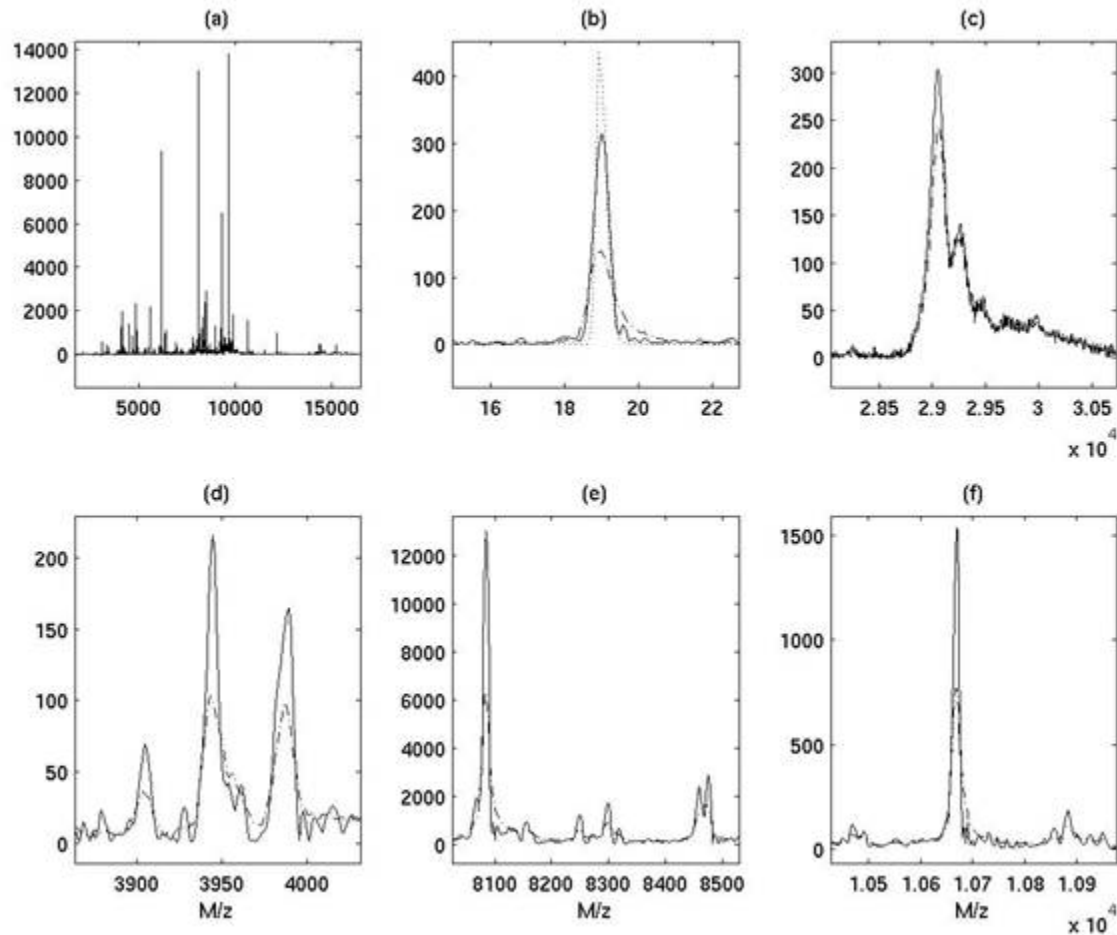
Putting it all together



Enhanced Resolution of Calibrant Peaks

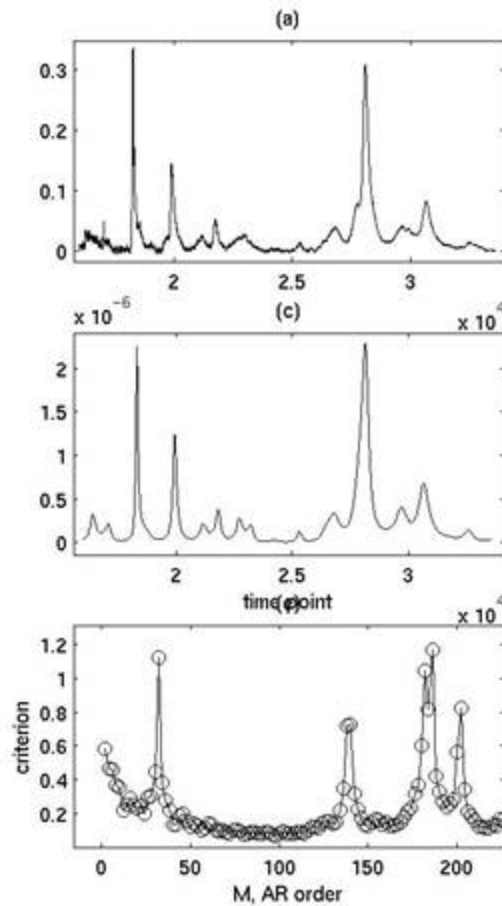


Enhanced Resolution in Pooled Serum



Default

BKG-Sub, MAV, Var-Rescale



Summary

Improving the processing of data output can dramatically improve sensitivity, resolution and reproducibility.

The Fold improvement may equal that of the “High resolution” SELDI-QStar.

Lookout for default Settings

Eastern Virginia Medical School Biomarker Discovery Laboratory

Investigators

John Semmes, Ph.D.
John Davis, M.D.
Jose Diaz, M.D., Ph.D.
Rick Drake, Ph.D.
Christine Laronga, M.D.
Paul Schellhammer, M.D.
Jeffery T. Wadsworth, M.D.

Fellows

Alberto Corica, M.D.
Daniel Holterman, Ph.D.
Gunjan Malik, Ph.D.
Lining Qi, Ph.D.

Staff

Diane Brassil
Lisa Cazares
MaryAnn Clements
Tarek Kendil
Brian Main
Michelle Moody
Michael Ward

E
V
M
S



Biostatistics/Computation

WMRI

William Cooke, Ph.D.
Dasha Malyrenko, Ph.D.
Denis Manos, Ph.D.
Michael Trossett, Ph.D.
Eugene Tracy, Ph.D.